

OptiPrep™ Mini-Review MS17

Resolution of soluble cytosolic proteins from membrane vesicles and organelles: a bibliography

There are three Application Sheets listed in the Application Sheet Index under “Protein localization (membrane *versus* cytosol)” which describe different gradient strategies; they can be found on the OptiPrep™ Applications flash drive or accessed via the following website www.axis-shield-density-gradient-media.com (click on “Methodology”, then “Organelles and subcellular membranes” and follow the links from the Index):

- ◆ **Discontinuous gradient: OptiPrep™ Application Sheet S35**
- ◆ **Self-generated gradient: OptiPrep™ Application Sheet S36**
- ◆ **A special strategy for rapid resolution of protein complexes and cytosol: OptiPrep™ Application Sheet S37**
- ◆ **Note that Mini-Reviews addressing the resolution of mammalian cell exosomes and other microvesicles from soluble proteins are covered in OptiPrep™ Mini-Review MS13 and the similar resolution of bacterial and fungal microvesicles in OptiPrep™ Mini-Review MS14.**

The reference list, which follows, includes principally papers describing the separation of membranes and soluble (cytosolic) proteins (**Section 1**); it is divided alphabetically into source material (**cell or tissue type**). It includes both mammalian and non-mammalian sources and in each of the 26 sections, references are listed alphabetically according to first author. **Section 2** lists a few papers that report the study of previously prepared subcellular membranes to determine the distribution of a particular protein between the soluble fraction and the organelle(s). Others describe the separation of vesicles either budded from the cells or obtained from permeabilized cells. In some cases gradients also resolve lipid droplets.

- ◆ Key words in titles are highlighted in light blue.

1. Cells or tissues

1.1. Algae

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1.2. Bacteria

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1.3. Brain

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1.11 Insect cells

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1.12. Jurkat cells

- Frankel, A.D.**, Alber, T., Zhou, Q. and Krogan, N.J. (2011) *Purification and characterization of HIV–human protein complexes* Methods, **53**, 13–19
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1.13. Kidney proximal tubule cells (incl. LLC-PK1)

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1.14. Liver

- Kanayama, M.**, Xu, S., Danzaki, K., Gibson, J.R., Inoue, M., Gregory, S.G. and Shinohara, M.L. (2017) *Skewing of the population balance of lymphoid and myeloid cells by secreted and intracellular osteopontin* Nat. Immunol., **18**, 973-984

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1.15 Lung cells

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1.16 MDCK cells

Grindstaff, K.K., Yeaman, C., Anandasabapathy, N., Hsu, S.C., et al (1998) *Sec6/8 complex is recruited to cell-cell contacts and specifies transport vesicle delivery to the basal-lateral membrane in epithelial cells* Cell, **93**, 731-740

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1.17. Monkey kidney cells

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1.18. Mouse embryo fibroblasts

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1.22 Plant cells

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1.24 Vero cells

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1.26 Yeast

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2. Subcellular membranes

2.1 Golgi membranes

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2.2 Lysosomes/endosomes

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2.3 Vesicles (budded and from permeabilized cells)

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