

# OptiPrep™ Reference List RC03

## Viable/non-viable cell separation

This Reference List includes, in Section 2, a short summary of the methodology

### 1. Introduction

Studies of cells from tissues (mammalian and non-mammalian) involve both mechanical and enzymic disaggregation of for example, liver or spleen or plant leaves, to release single cells. Prior to fractionation of different cell types it is often necessary to remove partially disrupted cells, released organelles and cytoplasm from damaged cells, other often fibrous debris from the tissue and any residual enzymes used in the disaggregation process.

### 2. Methodology

#### 2a. Sedimentation mode

In iodixanol solutions, non-viable cells are denser than viable cells

and there are two basic strategies for separating out the less dense viable cells, sedimentation or flotation. The often-used **Sedimentation Strategy 1**, is a simple density barrier (Figure 1). The density of the barrier is chosen to be slightly higher than that of the cells of interest. At the end of the centrifugation the viable cells band and non-viable cells separate across the barrier. The problems with this simple system are: (1) soluble proteins in the sample layer diffuse into the barrier; (2) while most nuclei will probably pellet, most subcellular particles released from disrupted cells will remain in sample layer zone; (3) partially disrupted cells will band across the viable cell layer. In **Sedimentation Strategy 2** (Figure 2) a lower density layer (1.05- 1.06 g/ml) is interposed between the resolving layer and the sample. The viable and non-viable cells are again separated across the denser layer but the diffusing soluble proteins will not reach the viable cell layer and there will be less contamination by smaller subcellular particles.

In both top-loading strategies all of the particles (cells, non-viable cells, partially broken cells, organelles) will be moving in the same direction and the problems described above will be compounded by the tendency for particles to aggregate at the interfaces. Hence it is usually necessary to wash the recovered viable cells several times to remove contamination by smaller particles and partially-disrupted cells.

#### 2b. Flotation mode (Strategy 3)

In the flotation mode the sample is adjusted to a density greater than that of the viable cells (Figure 3); this will allow the layering a lower density solution whose density is still greater than that of the viable cells. A small layer of saline is layered on top. During the centrifugation the viable cells float to the top interface; non-viable, partially broken cells and nuclei sediment slowly towards the bottom of the tube. The topmost layer (culture medium or saline) is not critical to the separation but prevents the viable cells from banding at an air/liquid interface. The big advantage of the method is that the viable cells move in the opposite direction to that of the non-viable and partially broken cells and nuclei. Aggregation at interfaces is therefore minimized and there should be layer of clean liquid that is essentially devoid of any contaminants.

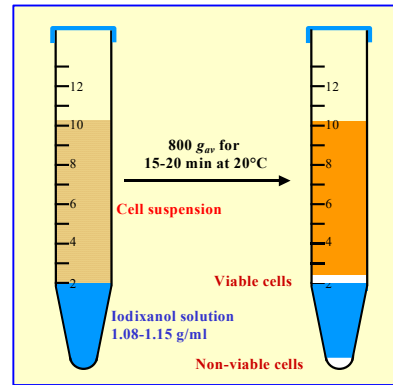


Figure 1: Sedimentation: Strategy 1

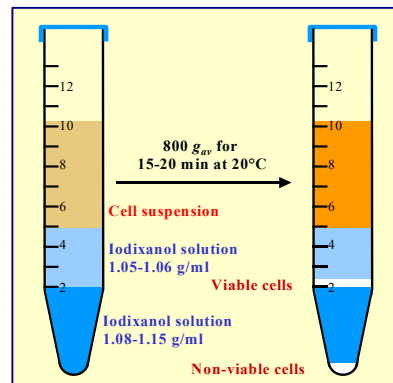


Figure 2: Sedimentation: Strategy 2

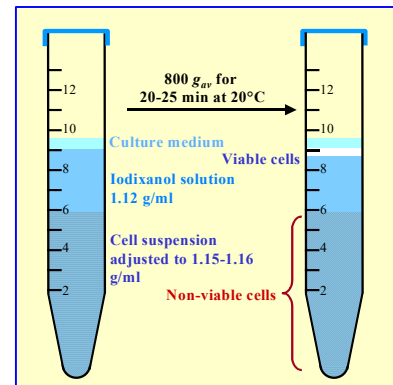


Figure 3: Flotation: Strategy 3

## 2c. Gradient solution preparation

All of the gradient solutions are very easily prepared from OptiPrep™, as sterile 60% (w/v) solution, which makes the preparation of gradient solutions very easy. The **methodology using iodixanol gradients for removal of non-viable cells** is given in **Application Sheet C14**. This is accessible from the following website: [www.OptiPrep.com](http://www.OptiPrep.com)

Note that although we recommend the OptiPrep™-based flotation strategy (outlined above and described in detail in **Application Sheet C14**), which should serve as a broadly applicable methodology, there are examples in the literature of the use of technical variations that may have a particular advantage, either for sample handling or for any subsequent technology. For example sedimentation on to a 1.10 g/ml barrier was used for melanoma cells [1,2]; the cell suspension was mixed with an equal volume of 24% (w/v) iodixanol (OptiPrep™ diluted with growth medium) and centrifuged at 800 g for 15 min to float viable epithelial cells [3]; HL60 cells were layered on top of three iodixanol solutions of density 1.057, 1.068 and 1.083 g/ml and centrifuged at 450 g for 30 min; the viable cells banded at the 1.057/1.068 g/ml interface [4]; a two-step iodixanol gradient (400 g for 15 min) was used for progenitor cells [5] and a plasma cell suspension was mixed 3.3:1 with OptiPrep™ (14% w/v iodixanol final concentration), centrifuged at 1400 g for 5 min to float the viable cells [6].

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2. Quintana, E., Shackleton, M., Foster, H.R., Fullen, D.R., Sabel, M.S., Johnson, T.M. and Morrison, S.J. (2010) *Phenotypic heterogeneity among tumorigenic melanoma cells from patients that is reversible and not hierarchically organized* Cancer Cell, **18**, 510–523
3. Bruce, A.T., Sangha, N., Richmond, A., Johnson, K., Jones, S., Spencer, T. and Ludlow, J.W. (2010) *Use of iodixanol self-generated density gradients to enrich for viable urothelial cells from non-neurogenic and neurogenic bladder tissue* Tissue Eng., Part C Methods **16**, 33-40
4. Hauert, A.B., Martinelli, S., Marone, C. and Niggli, V. (2002) *Differentiated HL-60 cells are a valid model system for the analysis of human neutrophil migration and chemotaxis* Int. J. Biochem. Cell Biol., **34**, 838-8541282
5. Howell, O.W., Scharfman, H.E., Herzog, H., Sundstrom, L.E., Beck-Sickinger, A and Gray, W.P. (2003) *Neuropeptide Y is neuroproliferative for hippocampal precursor cells* J. Neurochem., **86**, 646-659
6. Chatterjee, M., Stuhmer, T., Herrmann, P., Bommert, K., Dorken, B. and Bargou, R.C. (2004) *Combined disruption of both the MEK/ERK and the IL-6R/STAT3 pathways is required to induce apoptosis of multiple myeloma cells in the presence of bone marrow stromal cells* Blood, **104**, 3712-3721

## 3. Reference list of papers reporting the use of OptiPrep™

### Amphibian cells

**Cox, T.C.** (1999) *Calcium and ATP regulation of ion transport in larval frog skin* J. Comp. Physiol. B **169**, 344-350

### Bacteria

**Dehusa, O.**, Pfitzenmaier, M., Stuebs, G., Fischer, N., Schwaeble, W., Morath, S., Hartung, T., Geyer, A. and Hermann, C. (2011) *Growth temperature-dependent expression of structural variants of Listeria monocytogenes lipoteichoic acid* Immunobiology, **216**, 24–31

### B cells

**Methot, S. P.**, Litzler, L.C., Subramani, P.G., Eranki, A.K., Fifield, H., Patenaude, A-M., Gilmore, J.C., Santiago, G.E., Bagci, H., Côté, J-F. et al (2018) *A licensing step links AID to transcription elongation for mutagenesis in B cells* Nat. Comm., **9**: 1248

### Bone marrow cells

**Khoramian Tusi, B.**, Wolock, S.L., Weinre, C., Hwang, Y., Hidalgo, D., Zilionis, R., Waisman, A., Huh, J.R., Klein, A.M. and Socolovsky, M. (2018) *Population snapshots predict early haematopoietic and erythroid hierarchies* Nature, **555**, 54-60

### Bronchoalveolar lavage cells

**Kotzin, J.J.**, Spencer, S.P., McCright, S.J., Uthaya Kumar, D.B., Collet, M.A., Mowel, W.K., Elliott, E.N., Uyar, A. et al (2016) *The long non-coding RNA Morrbid regulates Bim and short-lived myeloid cell lifespan* Nature, **537**, 239-243

#### Breast tumour/cancer cells

**Chopra, S.S.**, Jenney, A., Palmer, A., Niepel, M., Chung, M., Mills, C., Sivakumaren, S.C., Liu, Q., Chen, J-Y. (2020) *Torin2 exploits replication and checkpoint vulnerabilities to cause death of PI3K-activated triple-negative breast cancer cells* Cell Systems 10, 66–81

**Green, J.L.**, La, J., Yum, K.W., Desai, P., Rodewald, L-W., Zhang, X., Leblanc, M., Nusse, R., Lewis, M.T. and Wahl, G.M. (2013) *Paracrine Wnt signaling both promotes and inhibits human breast tumor growth* Proc. Natl. Acad. Sci. USA, **110**, 6991–6996

#### Cancerous tissue

**Zierhut, C.**, Yamaguchi, N., Paredes, M., Luo, J-D., Carroll, T. and Funabiki, H. (2019) *The cytoplasmic DNA sensor cGAS promotes mitotic cell death* Cell **178**, 302–315

#### Cardiomyocytes

**Shaikh, S.R.**, Dumauval, A.C., Castillo, A., LoCasio, D., Siddiqui, R.A., Stillwell, W. and Wassall, S.R. (2004) *Oleic and docosahexaenoic acid differentially phase separate from lipid raft molecules: a comparative NMR, DSC, AFM, and detergent extraction study* Biophys. J., **87**, 1752-1766

**Siddiqui, R.A.**, Shaikh, S.R., Kovacs, R., Stillwell, W. and Zaloga, G. (2004) *Inhibition of phenylephrine-induced cardiac hypertrophy by docosahexaenoic acid* J. Cell. Biochem., **92**, 1141-1159

**Wenigke, K.**, Debierre-Grockiego, F., Wichmann, D., Brattig, N.W., Pankuweit, S., Maisch, B., Schwarz, R.T. and Ruppert, V. (2008) *Glycosylphosphatidylinositol-induced cardiac myocyte death might contribute to the fatal outcome of Plasmodium falciparum malaria* Apoptosis, **13**, 857-866

**Wichmann, D.**, Schwarz, R.T., Ruppert, V., Ehrhardt, S., Cramer, J.P., Burchard, G.D., Maisch, B. and Debierre-Grickiego, F. (2007) *Plasmodium falciparum glycosylphosphatidylinositol induces limited apoptosis in liver and spleen mouse tissue* Apoptosis, **12**, 1037-1041

#### Colonic macrophages

**Filardy, A.A.**, He, J., Bennink, J., Yewdell, J. and Kelsall, B.L. (2016) *Posttranscriptional control of NLRP3 inflammasome activation in colonic macrophages* Mucosal Immunol., **9**, 850-858

#### Dendritic cells

**Kapadia, C.H.**, Tian, S., Perry, J.L., Sailer, D., Luft, C.J., DeSimone, J.M. (2018) *Extending antigen release from particulate vaccines results in enhanced antitumor immune response* J. Control. Release, **269**, 393–404

#### Electroporation-treated cells

**Steinbrunn, T.**, Chatterjee, M., Bargou, R.C. and Stühmer, T. (2014) *Efficient transient transfection of human multiple myeloma cells by electroporation – an appraisal* PLoS One, **9**: e97443

#### Epithelial cells

**Bruce, A.T.**, Sangha, N., Richmond, A., Johnson, K., Jones, S., Spencer, T. and Ludlow, J.W. (2010) *Use of iodixanol self-generated density gradients to enrich for viable urothelial cells from non-neurogenic and neurogenic bladder tissue* Tissue Eng., Part C Methods **16**, 33-40

#### Hepatocytes (see also “Human foetal liver cells”)

**Schmelzer, E.**, Wauther, E. and Reid, L.M. (2006) *The phenotypes of pluripotent human hepatic progenitors* Stem Cell., **24**, 1852-1858

**Sicklick, J.K.**, Li, Y-X., Melhem, A., Schmelzer, E., Zdanowicz, M., Huang, J., Caballero, M., Fair, J.H., Ludlow, J.W., McLelland, R.E., Reid, L.M. and Diehl, A.M. (2006) *Hedgehog signaling maintains resident hepatic progenitors throughout life* Am. J. Physiol. Gastrointest. Liver Physiol., **290**, G859-G870

#### HL-60 cells

**Hauert, A.B.**, Martinelli, S., Marone, C. and Niggli, V. (2002) *Differentiated HL-60 cells are a valid model system for the analysis of human neutrophil migration and chemotaxis* Int. J. Biochem. Cell Biol., **34**, 838-854/282

#### Human foetal liver cells

**Khoramian Tusi, B.**, Wolock, S.L., Weinre, C., Hwang, Y., Hidalgo, D., Zilionis, R., Waisman, A., Huh, J.R., Klein, A.M. and Socolovsky, M. (2018) *Population snapshots predict early haematopoietic and erythroid hierarchies* Nature, **555**, 54-60

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**Schmelzer, E.** and Reid, L.M. (2009) *Human telomerase activity, telomerase and telomeric template expression in hepatic stem cells and in livers from fetal and postnatal donors* Eur. J. Gastroenterol. Hepatol., **21**, 1191-1198

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**Pan, C.W.**, Horvath, D.G., Braza, S., Moore, T., Lynch, A., Feit, C. and Abbyad, P. (2019) *Sorting by interfacial tension (SIFT): label-free selection of live cells based on single-cell metabolism* Lab Chip, **19**, 1344-1351

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**Quintana E.**, Shackleton, M., Sabel, M.S., Fullen, D.R., Johnson, T.M. and Morrison, S.J. (2008) *Efficient tumour formation by single human melanoma cells* Nature **456**, 593-599

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**Chatterjee, M.**, Stuhmer, T., Herrmann, P., Bommert, K., Dorken, B. and Bargou, R.C. (2004) *Combined disruption of both the MEK/ERK and the IL-6R/STAT3 pathways is required to induce apoptosis of multiple myeloma cells in the presence of bone marrow stromal cells* Blood, **104**, 3712-3721

**Olsen, O.E.**, Sankar, M., Elsaadi, S., Hella, H., Buene, G., Darvekar, S.R., Misund, K., Katagiri, T., Knaus, P. and Holien, T. (2018) *BMPR2 inhibits activin and BMP signaling via wild-type ALK2* J. Cell Sci., 131: jcs213512

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**Vätsveen, T.K.**, Børset, M., Dikic, A., Tian, E., Micci, F., Lid, A.H.B., Meza-Zepeda, L.A., Coward, E. et al (2016) *VOLIN and KJON-Two novel hyperdiploid myeloma cell lines* Genes Chromosomes Cancer, **55**, 890-901

**Zöllinger, A.**, Stühmer, T., Chatterjee, M., Gattenlöhner, S., Haralambieva, E., Müller-Hermelink, H-K., Andrusis, M., Greiner, A., Wesemeier, C., Rath, J.C., Einsele, H. and Bargou, R.C. (2008) *Combined functional and molecular analysis of tumor cell signaling defines 2 distinct myeloma subgroups: Akt-dependent and Akt-independent multiple myeloma* Blood, **112**, 3403-3411

#### Myoblasts/myocytes

**Benabdallah, B.F.**, Bouchentouf, M. and Tremblay, J.P. (2005) *Improved success of myoblast transplantation in mdx mice by blocking the myostatin signal* Transplantation, **79**, 1696-1702

Liu, S.J. (2013) *Characterization of functional capacity of adult ventricular myocytes in long-term culture* Int. J. Cardiol., **168**, 1923-1936

#### Pancreatic acinar cells

**Mankad, P.**, James, A., Siriwardena, A.K., Elliott, A.C. and Bruce, J.I.E. (2012) *Insulin protects pancreatic acinar cells from cytosolic calcium overload and inhibition of plasma membrane calcium pump* J. Biol. Chem., **287**, 1823-1836

#### Progenitor cells

**Howell, O.W.**, Scharfman, H.E., Herzog, H., Sundstrom, L.E., Beck-Sickinger, A and Gray, W.P. (2003) *Neuropeptide Y is neuroproliferative for hippocampal precursor cells* J. Neurochem., **86**, 646-659

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#### Spinal cells

**Nguyen, H.X.**, Galvan, M.D. and Anderson, A.J. (2008) *Characterization of early and terminal complement proteins associated with polymorphonuclear leukocytes in vitro and in vivo after spinal injury* J. Neuroinflamm., **5**:26

#### Spleen cells

**Deloizy, C.**, Fossum, E., Barnier-Quer, C., Urien, C., Chrun, T., Duval, A., Codjovi, M., Bouguyon, E., Maisonnasse, P. et al (2017) *The anti-influenza M2e antibody response is promoted by XCR1 targeting in pig skin* Sci. Rep., **7**: 7639

**Westhorpe, C.L.V.**, Norman, M.U., Hall, P., Snelgrove, S.L., Finsterbusch, M., Li, A., Lo, C., Tan, Z.H. et al (2018) *Effector CD4+ T cells recognize intravascular antigen presented by patrolling monocytes* Nat. Comm., **9**: 747

#### T-cells

Westhorpe, C.L.V., Norman, M.U., Hall, P., Snelgrove, S.L., Finsterbusch, M., Li, A., Lo, C., Tan, Z.H. et al (2018) *Effector CD4+ T cells recognize intravascular antigen presented by patrolling monocytes* Nat. Comm., **9**: 747

#### Urological cancer cells

**Oates, J.E.**, Grey, B.R., Addla, S.K., Samuel, J.D., Hart, C.A., Ramani, V.A.C., Brown, M.D. and Clarke, N.W. (2009) *Hoechst 33342 side population identification is a conserved and unified mechanism in urological cancers* Stem Cell Devel., **18**, 1515-1521

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